TOTAL SYNTHESIS OF LEUKOTRIENE \mathbf{B}_{5}

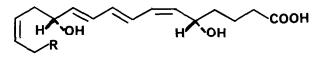
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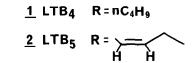
Abstract: A practical, stereocontrolled synthesis of leukotriene B₅ is described which makes this substance readily available for the first time.

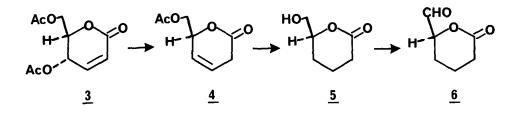
Previous papers from this laboratory have described two total syntheses of leukotriene B_{A} (1) which served the dual purpose of elucidating the stereochemistry of the conjugated triene unit and providing adequate material for biological study.^{1,2} The analog of 1 which originates from eicosapentaenoic acid rather than arachidonic acid, LTB_5 (2), is of interest in connection with understanding the basis for the cardiovascular protective effect of dietary fish lipids.³ Previously LTB₅ has only been obtained in minute amount (a few micrograms)⁴ from biosynthetic experiments, and the study of its biological effects has been severely limited in consequence. We report here the first total synthesis of LTB₅ by a practical process which is stereocontrolled and does not require resolution.

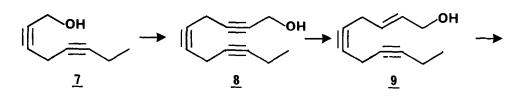
The known lactone 3,⁵ which is readily available from commercial tri-O-acetyl-D-glucal,⁶ was converted to the aldehyde $\underline{6}$ by the following sequence: (1) reduction of $\underline{3}$ (10 equiv. of zinc am algum, anhydrous hydrogen chloride, ether, 0°, 2 hr., 90% yield) to give $\frac{4}{2} ([\alpha]_D^{20} -113.2^\circ (c \ 1.25, CHCl_2));$ (2) base-catalyzed double bond transposition (0.1 equiv. diazobicyclo[5.4.0]undecene, tetrahydrofuran (THF), 25°, 1 hr., 99% yield); (3) base hydrolysis (1M lithium hydroxide, dimethoxyethane (DME), 1:1, 25°, 2 hr.); (4) hydrogenation (1 atm.) over 10% Pd-C in ethyl acetate to give $5 ([\alpha]_D^{20} + 34.68^\circ)$ (c 1.3, CHCl₂), 75% yield from 4); and (5) oxidation at 23° for 1.5 hr. with pyridinium dichromate⁷ (5 equiv., anhydrous magnesium sulfate, powdered 4Å molecular sieves, CH₂Cl₂, 78%).

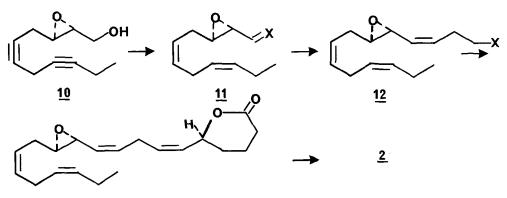
Octa-2, 5-diyn-1-ol $\chi^{8,9}$ was converted to the triphenylphosphonium iodide 12 (X= $\dot{P} \dot{\Phi}_{3} I$) by the following sequence: (1) primary iodide formation¹⁰ with triphenylphosphine, imidazole and iodine (1.5 equiv. of each reagent, ether-acetonitrile (CH₃CN), 3 : 1, 25°, 40 min., 97%); (2) copper (I) catalyzed alkylation with the bisbromom agnesium salt of propargyl alcohol¹¹ (5 equiv., THF, 65°, 12 hr., $7 \longrightarrow 8$, 90%); (3) reduction with lithium aluminum hydride¹² in ether at 25° for 5 hr. (8 \longrightarrow 9, 91%); (4) enantioselective epoxidation¹³ using dimethyl (S,S) (-) tartrate (9, \rightarrow 10, 76%, 10: $[\alpha]_{D}^{23}$ +8.3° (c 2.35, CHCl₃)); (5) hydrogenation (1 atm.) over Lindlar catalyst deactivated with triethylamine in THF $(10 \rightarrow 11)$ (X= H, OH), 90%, 11 (X= H, OH): $[\alpha]_D^{23}$ + 20.5° (c 1.73, CHCl₃)); (6) oxidation with Collins reagent (6 equiv., $11 (X=H, OH) \rightarrow 11 (X=O)$, 84%; (7) exposure of 11 (X=O) to 1.25 equivalents of the ylide prepared from the 2-methoxy-2-propyl ether of (3-hydroxypropyl)triphenylphosphonium













bromide¹⁴ (using sodium hexamethyldisilazide,¹⁵ THF-toluene, 1 : 2, 0°, 20 min.) at -78° for 10 min. and from -78° to 0° over 1 hr. $(11 (X = O) \rightarrow 12 (X = OC (OCH_3)(CH_3)_2, 94\%);$ (8) acidic hydrolysis (acetic acid-water-CH₃CN, 0.006 : 1 : 4, 25°, 6 hr., 12 (X = OC (OCH₃)(CH₃)₂ \rightarrow 12 (X = OH), 84%); (9) primary iodide formation with triphenylphosphine, imidazole, iodine, dlisopropylamine (1.5 equiv. of each reagent)¹⁰ (ether - CH₃CN, 3 : 1, 25°, 40 min., 12 (X = OH) \rightarrow 12 (X = I); and (10) displacement of iodide with triphenylphosphine (1.5 M, 3 equiv., CH₃CN, 60°, 16 hr., 12 (X = I) \rightarrow 12 (X = $\overrightarrow{P} \phi_3 \overrightarrow{I}$), 92% from 12 (X = OH)).

The phosphonium iodide 12 (X = $P \Phi_3 I$) was converted to the corresponding ylide (0.95 equiv. of nBuLi, THF, -78°, 1.5 hr.) and treated sequentially with hexamethylphosphoric triamide (15 equiv.) and then the aldehyde <u>6</u> (1.2 equiv.). After 30 min. at -78° the reaction mixture was warmed to 0° over 1.5 hr. Extractive isolation and thin-layer chromatography gave the lactone 13 (45%) and the less mobile 6,7-<u>trans</u> isomer (5%) of 13. Exposure of 13 to potassium isopropoxide in isopropanol (10 equiv., 0.5 <u>M</u>, 0°, 1 hr.) followed by basic hydrolysis afforded LTB₅ (2) in 75% yield and >95% purity by RP-HPLC analysis. Both RP-HPLC retention volumes¹⁶ (Dupont Zorbax ODS, 4.6mm x 25 cm, CH₃OH-H₂O-HOAc, 60 : 40 : 0.1, 2.0 ml/min., retention volume 21.5) and the UV spectra (UV max (CH₃OH) 260.2, 269.3, 280.3 nm) of biosynthetic LTB₅ (2)¹⁷ and synthetic 2 were identical. The synthetic LTB₅ (2) was fully characterized as its methyl ester diacetate by HPLC, ¹⁸ ¹H NMR, ¹⁹ UV, ²⁰ and MS²¹ analysis.

The availability of synthetic LTB_5 allowed comparative studies of its biological action on human polymorphonuclear leukocytes relative to LTB_4 . Synthetic LTB_5 was found to be only <u>ca</u>. 5% as active as LTB_4 in leukocyte chemotaxis and aggregation.²² LTC_4 and LTC_5 , in contrast, appear to be similar in spasmogenic activity.²³ It is possible that part of the cardioprotective effect of fish lipid may be associated with the diminished activity of LTB_5 toward neutrophils relative to LTB_4 .²⁴

References and Notes

- 1. E. J. Corey, A. Marfat, G. Goto, and F. Brion, J. Am. Chem. Soc., 102, 7984 (1980).
- 2. E. J. Corey, A. Marfat, K. S. Kim, P. B. Hopkins, and F. Brion, <u>Tetrahedron Letters</u>, <u>22</u>, 1077 (1981).
- 3. For a recent summary of this literature see, E. J. Corey, C. Shih, and J. R. Cashman, <u>Proc.</u> <u>Natl. Acad. Sci. USA</u>, <u>80</u>, 3581 (1983).
- 4. R. A. Murphy, W. C. Pickett, Brenda R. Culp, and W. R. Lands, Prostaglandins, 22, 613 (1981).
- J. Mieczkowski, J. Jurczak, M. Chmielewski, and A. Zamojski, <u>Carbohydrate Research</u>, <u>56</u>, 180 (1977).
- 6. P. Jarglis and F. W. Lichtenthaler, <u>Tetrahedron Letters</u>, <u>23</u>, 3781 (1982).
- 7. E. J. Corey and G. Schmidt, Tetrahedron Letters, 399 (1979).
- 8. S. S. Nigam and B. C. L. Weedon, J. Chem. Soc., 4049 (1956).
- Prepared from pent-2-yn-1-ol by conversion to the iodide¹⁰ (Pφ₃, I₂, imidazole, 1.5 equiv. of each reagent, ether-CH₃CN, 3 : 1, 25°, 40 min.) and then Cu(I) catalyzed alkylation with the bisbromo-magnesium salt of propargyl alcohol¹¹ in 98% overall yield.
- 10. P. J. Garegg and B. Samuelsson, J. Chem. Soc., Perkin I, 2866 (1980).
- 11. J. M. Osbond, R. G. Philpott and J. C. Wickens, J. Chem. Soc., 2779 (1961).
- 12. E. J. Corey, A. Marfat and G. Goto, J. Am. Chem. Soc., 102, 6607 (1980).
- 13. T. Katsuki and K. B. Sharpless, J. Am. Chem. Soc., <u>102</u>, 5974 (1980).

- 14. E. J. Corey, P. B. Hopkins, J. E. Munroe, A. Marfat and S.-i. Hashimoto, <u>J. Am. Chem. Soc.</u>, 102, 7986 (1980).
- 15. H. Bestman, W. Stransky and O. Vostrowski, Chem. Ber., 109, 1694 (1976).
- 16. RP-HPLC retention volumes for LTB₅ (2) and LTB₄ (1) were 8.6 and 12.6, respectively (Dupont Zorbax ODS, 4.6 mm x 25 cm., CH₃OH-H₂O-HOAc, 75 : 25 : 0.01, 2.0 ml./min.); cf. ref. 4.
- 17. We are grateful to Dr. Tak Lee of the Harvard Medical School for supplying biosynthetic $LTB_e(2)$.
- Dupont Zorbax Sil 4.6 mm x 25 cm.; 0.3% iPrOH 3% EtOAc hexane; 2.0 ml./min., retention volume 16.
- 19. 1 H NMR (d⁶-benzene, 270 MHz): 6.79 (dd, J = 11.53, 14.83 Hz, H 8); 6.34 (dd, J = 10.55, 14.83 Hz, H 10); 6.09 (dd, J = 10.87, 14.83 Hz, H 9); 6.03 (t, J = 11.53 Hz, H 7); 5.90 (dt, J = <u>ca</u>. 9.5, 11 Hz, H 5); 5.63 (dd, J = 6.92, 14.82 Hz, H 11); 5.60 5.38 (m, 5H, H 12, H 14, H 15, H 17, H 18); 5.33 (t, J = 10.22 Hz, H 6); 3.35 (s, 3H); 2.83 (m, 2H, H 16), 2.47 (m, 1H, H 13), 2.38 (m, 1H, H 13); 1.74 (s, 3H); 1.68 (s, 3H); 0.95 (t, J = 7.5 Hz, 3H, H 20). This spectrum was consistent with that of the methyl ester diacetate of LTB₄ (1), E. J. Corey, P. B. Hopkins, A. E. Barton, B. Bangerter, and P. Borgeat, <u>Tetrahedron</u>, <u>38</u>, 2653 (1982).
- 20. UV_{max} (CH₃OH) 260.2, 269.3, 280.0 nm.
- 21. MS (CI, NH₃, 320°) 450 (M + NH₄⁺, 100%), 392 (M + NH₄⁺ HOAc, 46%), 332 (M + NH₄⁺ 2 (HOAc), 53%).
- 22. T. H. Lee, J.-M. Huerta, C. Shih, S. G. Pyne, E. J. Corey, R. A. Lewis, and K. F. Austen, J. Biol. Chem., in press.
- 23. S. Hammarstrom, J. Biol. Chem., 255, 7093 (1980).
- 24. This research was assisted financially by grants from the National Science Foundation and the National Institutes of Health.

(Received in USA 18 July 1983)